

ASSESSMENT OF BIOACTIVE COMPOUNDS PRODUCED BY ENDOPHYTIC FUNGUS ISOLATED FROM *SCLEROCARYA BIRREA* PLANT

*¹Garba, S. A., ²Mudi, S.Y., ³Muhammad, A. H., ⁴Abdullahi, S.

¹Chemistry Department, Faculty of Natural and Applied Science, Sule Lamido University, Kafin Hausa, Nigeria

²Department of Pure and Industrial Chemistry, Bayero University, Kano

³Department of Biological Sciences, Sule Lamido University, Kafin Hausa

⁴National Biotechnology Development Agency, Abuja

*Corresponding Author: sulaimanabubakar.garba@jsu.edu.ng

ABSTRACT

This research was aimed at the characterization of bioactive components of endophytic fungi isolated from the stem bark of *Sclerocarya birrea* plant using NMR, GC-MS and FT-IR. The stem of this plant was collected, prepared and screened for endophytic fungi. The isolated endophytic fungi were fermented for large scale production on potato dextrose broth and extracted with ethyl acetate. This crude extract was fractionated and purified over silica-gel column chromatography. The fractions obtained were tested for antimicrobial activity and most active isolate (SB7-FO-F₁₀) were characterized. Isolate SB7-FO-F₁₀ showed antibacterial activity with zone of inhibition of 13mm, 14mm and 14mm against *E. coli*, *Streptococcus pyogenes* and *Pseudomonas aeruginosa* respectively. From the spectral analyses, isolate SB7-FO-F₁₀ afforded a compound characterized to be 6-(5-ethoxypentyl) 1-pentyl-2-methylhex-2-enedioate.

Key words: Bioactive, Characterization, Column Chromatography, Endophytic Fungi.

INTRODUCTION

Natural products, because of their ecological safety and greater variety of chemical structures compared to synthetic compounds, are regarded as powerful, promising source of novel compounds as well as hopeful choice for innovation in both medicine and agro chemistry. Plant is the chief source of natural products, while those from microorganisms tend to be more bioactive than the synthetic ones [1, 2]. Microorganisms are underutilized sources of natural products. They evolve in biosynthesis pathway and mechanism for synthesizing a rich arsenal of complex secondary metabolites, particularly those from bacteria and fungi. Very little of them are known [3, 4].

Sclerocarya birrea is an African medicinal plant used to cure diseases and heal injuries. The larval stage of the beautiful green African moth *Argemamimosae* feeds on marula leaves [5-7]. In Nigeria and some other African countries, the stem bark, roots and leaves of *S. birrea* are used for an array of human ailments, including malaria fever, diarrhoea, dysentery, stomach ailments, headache, toothache and body pains [8]. Anticonvulsant effect of aqueous stem bark of *S. birrea* extract in mice was reported by Ojewale *et al* [9].

An endophytic fungus has been described as fungi that asymptotically colonize healthy plant tissues, even though they may after incubation or a latency period cause disease [10]. Several endophytes secrete secondary metabolites that protect plant against insect, pests, pathogenetic organism and herbivores [11]. They proved to be the promising sources of biologically active product of interest for specific medicinal applications [12]. Researches toward identification of bioactive metabolites lead to discovery of many new active compounds from different types of endophytic fungi such as podophyllotoxin, a well-known aryl tetralin lignan with potent anticancer, antiviral, antioxidant, antibacterial, immunostimulation and anti-rheumatic properties [13-19] and anti-fungal agents, oxysporidinone [20] and 6-epi-oxysporidinone [21], which were isolated from the *Fusarium* genus. This present research was aimed to screen, identify the potential novel antibiotic-producing endophytic fungi from the tissue of *Sclerocarya birrea* plant and characterize the isolated bioactive component using NMR, GC-MS and FT-IR

MATERIAL AND METHOD

Plant Sampling and Isolation of Endophytic Fungi

The plant samples were collected from Botanical and Ecological Gardens, Department of Botany, Bayero University, Kano (Nigeria). The plant surface was sterilized according to [22-24] and the sterilized segments were placed in a petri dish containing the potato dextrose agar (PDA) medium with chloramphenicol 280 mgml⁻¹ to suppress bacterium contamination. After an incubation of seven days, new fungal colonies were monitored and picked out. Individual fungal colonies were picked out and sub-cultured in PDA and the isolated fungi were stored at low temperature.

Identification of Fungi

The fungal isolates were prepared on slides and stained with lactophenol cotton blue reagent and examined with bright-field and phase-contrast microscope [24].

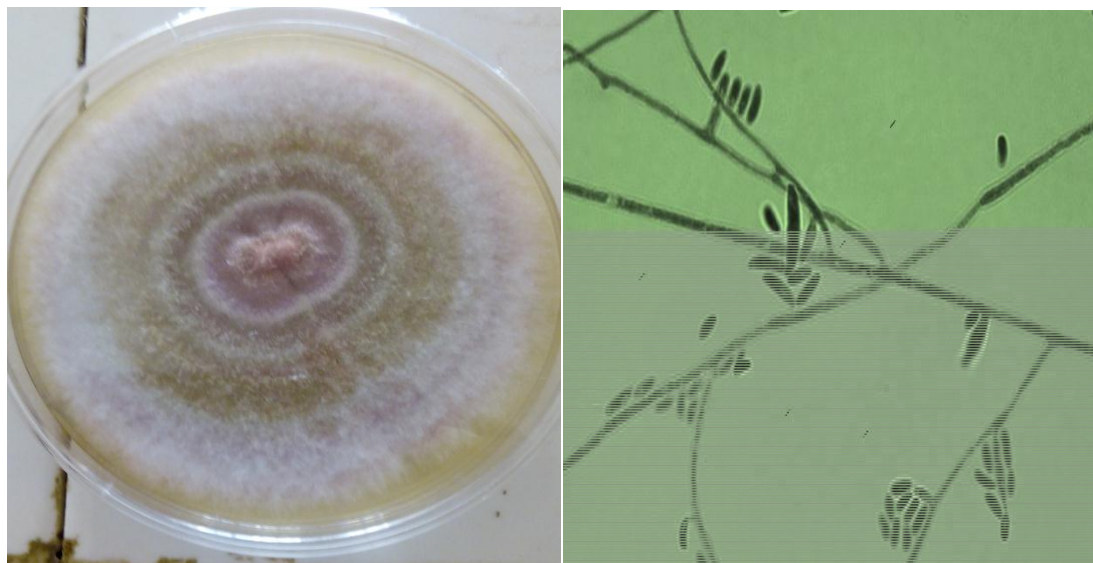


Plate 1: *Fusarium oxysporum*

Cultivation for Screening of Secondary Metabolites

About 100 mL of the potato dextrose broth was dispensed in 250 mL conical flask each and autoclave at 121 °C, 15lb for 15 min. Plug of the fungal material of about 2cm x 2cm were cut into each flask covered with bunk and kept static for about 30 days at room temperature. Flasks were regularly observed for fungal purity and similarities while variations were considered as contaminated and were discarded.

Extraction of the secondary metabolites from *Fuseriumoxysporum*

The mycelium was separated from the liquid by suction filtration using vacuum pump and the fungal mycelium was soaked in methanol prior to extraction, while the broth filtrate was extracted with ethyl acetate thrice. The broth ethyl acetate was partitioned with brine solution to remove unwanted materials. It was afterwards separated from the brine solution and dried with anhydrous MgSO₄ and finally concentrated at reduced pressure. The mycelia were soaked in methanol thrice for one week each and filtered by suction to afford the methanolic extracts which was concentrated at reduced pressure to yield orange-red residue. The methanolic crude extract was further partitioned between water and ethyl acetate (1:1) to yield the ethyl acetate fraction which was washed with brine solution and dried with anhydrous MgSO₄ and finally concentrated at reduced pressure to yield an orange-red solid 11.1g which was labeled as **SB7-FO**.



Plate 1: Mycelium of *F. oxysporum* grown on Potato Dextrose Broth

Purification of SBO-FO Ethyl Acetate Extraction

The ethyl acetate crude extract (5.0 g) was subjected to silica gel column chromatography in a glass column (121 x 2.5cm capacity), eluting with a total volume of 500 mL of hexane-chloroform gradient (100:0.00; 1:1; 2:3; 1:4; and 0.00:100), followed by chloroform: ethyl acetate, 9: 1; 4:1; 3:2; 1:1; 2:3; 1:4; 100% ethyl acetate and finally washed with acetone. About 50 mL of eluent was collected at intervals. The entire column produced 120 fractions. These fractions were pooled to give nineteen fractions which were later pooled to give three fractions.

Purification of F67 of SB7-FO

Fraction obtained from F₆₇ of SB7- FO samples was further purified over silica gel column chromatography eluting total volume of 100 mL hexane (100 %), hexane: chloroform (60:40; 50:50; 40:60; 100 % chloroform), chloroform: ethyl acetate (60:40; 50:5; 40:60; 100% ethyl acetate) and finally washed with 100 % acetone. About 20 mL eluent was collected at an interval which resulted in 50 fractions. F₂₉ obtained from the main column of F₆₇ (SB7-FO) sample was further purified to give eleven fractions after pooling those with the same R_f value and code with SB7-FO-F_n where n is the fraction number.

Antibacterial assay

To assess the antimicrobial activity of the endophytic fungi, five clinically isolated *Escherichia coli*, *Klebsiella pneumoniae*, *Streptococcus pyogenes*, *Staphylococcus aureus* and *Pseudomonas aeruginosa*, strains were obtained from Microbiology Department, Bayero University, Kano. Two loop fills of the standard inoculum were steadily streaked onto the prepared nutrient agar plates and discs of different concentrations as well as amoxicillin which served as the positive control were aseptically placed and pressed firmly onto each plate using sterile forceps at about 40 mm apart. After 15 min of free diffusion time, the plates were incubated at 37 °C for 24 h. Diameters of zones of inhibition were measured using a millimeter rule and recorded to the nearest whole number.

RESULTS AND DISCUSSION

During the screening of endophytic fungi from the tissue of *S. birrea* plant, three different species of *Fusarium* were isolated. The potential novel antibiotic-producing endophytic fungi were identified for exact genus according to its morphological features and conidia as *F. oxysporum*. Based on preliminary antimicrobial study, about sixty liters of *F. oxysporum* were fermented on potato dextrose broth for a period of four months to yield 11.7 g of crude ethyl acetate extract which was later subjected to purification over silica-gel column chromatography. Fractions from fungal strain SB7-FO showed only antibacterial activity (Table 1), even though their action against the microorganisms used was selective. SB7-FO-F₅, SB7-FO-F₁₀ and SB7-FO-F₃₀ were found to show potent antibacterial activity, with SB7-FO-F₁₀ been more effective against *P. aeruginosa* with zone of inhibition of 14 mm at 1000µg/mL followed by 13 mm and 14 mm at 1000µg/mL for *S. pyogenes* and *E.coli* respectively. SB7-FO-F₃₀ was more effective on *S. aureus* with zone of inhibition of 12 mm at 1000µg/mL followed by *S. pyogenes* with 14mm at 1000µg/mL, then *E.coli* with zone inhibition of 10 mm at 1000µg/mL and SB7-FO-F₅ was more effective against *S. pyogenes*, *S. aureus*, *K. pneumoniae* and *P. aeruginosa* with inhibition of 14 mm, 11 mm, 12 mm and 10 mm at 1000µg/mL respectively. The effect of these fractions was average when compared with the effect of the drug as control which has a maximum zone of inhibition of 26mm at 10µg/disc against *K. pneumoniae*.

The potential endophytic fungi have been identified for the exact genus based on morphological colony and spore or conidia characteristic as *F. oxysporum*. The compound isolated in this study

was characterized as a fatty acid ester 6-(5-ethoxypentyl) 1-pentyl-2-methylhex-2-enedioate and possess a potential biological activity.

Table 1: Antibacterial Activity of Column Fractions (F₂₉ of F₆₇) from the Crude Extract of SB7-FO

Sample	Concentration (µg/disc)	Zone of Inhibition (mm)				
		<i>E. coli</i>	<i>S. pyogenes</i>	<i>S. aureus</i>	<i>K. pneumoniae</i>	<i>P. aeruginosa</i>
SB7-FO-F ₅	10	08	10	07	08	08
	100	08	10	07	08	10
	1000	09	14	11	12	10
SB4-FO-F ₁₀	10	08	07	08	08	09
	100	10	10	09	10	09
	1000	13	13	11	12	14
SB7-FO-F ₁₅	10	NA	07	NA	NA	NA
	100	NA	07	NA	NA	NA
	1000	NA	10	NA	NA	NA
SB7-FO-F ₂₃	10	NA	NA	NA	NA	NA
	100	NA	NA	NA	NA	NA
	1000	NA	NA	NA	NA	NA
SB7-FO-F ₂₄	10	08	NA	NA	07	NA
	100	10	NA	NA	08	07
	1000	10	NA	NA	08	07
SB7-FO-F ₃₀	10	07	07	10	07	07
	100	08	10	10	09	08
	1000	10	14	12	09	12
SB7-FO-F ₃₅	10	NA	NA	NA	NA	NA
	100	NA	NA	NA	NA	NA
	1000	NA	NA	NA	NA	NA
N mgw	10	NA	NA	NA	NA	NA
	100	NA	NA	NA	NA	NA
	1000	NA	NA	NA	NA	NA
SB7-FO-F ₄₅	10	NA	07	NA	07	NA
	100	07	NA	08	08	NA

	1000	07	NA	08	08	NA
Amoxicillin	10	25	19	26	24	26

Keys: **NA** = No Activity means zone of inhibition equal to 6mm. (+) = Zone of inhibition greater than 6mm. *E. coli*= *Escherichia coli*, *S. pyogenes*= *streptococcus pyogenes*, *S. aureus*= *Staphylococcus aureus*, *P. aeruginosa*= *Pseudomonas aeruginosa*, *K. pneumoniae*= *Klebsiella pneumoniae*.

Previous researches revealed that fatty acids and their esters exhibited strong antimicrobial activity against microorganisms, demonstrating some specificity for individual microbial species [25] and another report by Rahmani and Heydarian [26] which indicated that fatty acid ester are active metabolites in *Salicornia* species. The compound isolated in this work was found to possess potent antimicrobial activity against *S. pyogenes* and *P. aeruginosa* followed by *S. aureus* and *E. coli*. This is in agreement with finding of Chandrasekharan *et al* [27] that studied antimicrobial activity of fatty acid ester extracts of four halophytic plants, *Arthrocremum indicum*, *Salicornia brachiata*, *Suaedamaritima* and *Suaedamonoica* belonging to the family chenopodiaceae. Their composition was analyzed by GC-MS and the result was promising. Compounds of this type (fatty acid esters) were found to possess various types of biological activities as reported by Gehan *et al* [28] which stated that a similar compound, octadecanoic acid methyl ester possessed the ability to inhibit the growth of pathogenic bacteria and fungi. 10, 13-eicosadienoic acid methyl ester has been reported to possess excellent antibacterial and antifungal activities [29].

This present study is in agreement with reports of Huda [30] which reported the antimicrobial activity *F. oxysporum*. It also agrees with the finding of Abubakar *et al* [31] who reported that the stem bark of *Sclerocarya birrea* is used as traditional medicine in Kano metropolis. It is also supported by the statement of Muhammad *et al* [29] about the antimicrobial activity of some fatty acid esters isolated from *Nigella sativa* and that of Saikkonen *et al* [11] which stated that plants and their associated endophytes were found to produce the same natural compounds. This research supported few researchers [32-35] who reported the antimicrobial activities of biosynthesized compounds by plant endophytes.

CONCLUSION

The stem bark of *Sclerocarya birrea* plant was screened for endophytic fungi and four different species were isolated. Strain of two species was found to possess potent antimicrobial activity. The column chromatograph of crude extracts of these strains produced fractions that possess significant antimicrobial activity with some showing selectivity against the tested pathogens. A bioactive compound which was found to possess a significant anti antibacterial effect against the tested organisms was isolated from the crude ethyl acetate extract of *F. oxysporum* and characterized using NMR (^1H , ^{13}C), GC-MS and FT-IR as 6-(5-ethoxypentyl) 1-pentyl-2-methylhex-2-enedioate.

RECOMMENDATION

Further research using advance analytic techniques such as 2-D or 3-D NMR, high resolution GC-MS is recommended to elucidate and propose the exact structure of the isolated compound

REFERENCES

1. Schulz, B., Boyle, C., Draeger, S., Römmert, A.-K., Krohn, K. (2002) Endophytic fungi: a source of novel biologically active secondary metabolites. *Mycologicalresearch*, 106, 996–1004.
2. Tenguria, R. K., Khan., F. N., Quereshi, S., (2011).Endophytes - Mines of pharmacological therapeutics. *World Journal of Science and Technology*, 1(5), 127–149.
3. Newman, D. J., Cragg, G.M., Snader, K. M (2003). Natural products as sources of new drugs over the period 1981–2002, *Journal of Natural Products*, 66, 1022–1037
4. Kharwar, R.N., Mishra, A., Gond, S.K. Stierle, A. & Stierle, D. (2011). Anticancer compounds derived from fungal endophytes: their importance and future challenges. *Nat Prod Rep*, 26-1208
5. Van Wyk, B-E., Van Oudtshoorn, B. & Gericke, N (1997). *Medicinal plants of South Africa*. Briza Publications, Pretoria
6. Venter, F. & Venter, J-A. (1996). Making the most of indigenous trees. *Briza Publications, Pretoria*
7. Coates Palgrave, k. (1983). *Trees of Southern Africa*, 2nd Edition. Struik, Cape Town.
8. Masoko, P, Mmushi, T.J, Mogashoa, M.M, Mokgotho, M.P, Mampuru, L. J.& Howard, R.L

- (2008). In vitro evaluation of the antifungal activity of *Sclerocarya birrea* extracts against pathogenic yeasts. *African journal of biotechnology* 7 (20), 3521-3526. Retrieved from <http://www.academicjournals.org/AJB>
9. Ojewole, J.A., Mawoza, T. & Chiwororowo, O. (2010). *Sclerocarya Birrea* (A. Rich) Hochst; A review of its phytochemistry, pharmacology and toxicology its ethnomedicinal uses. *Phytother Res* 24(5), 633-9.
 10. Stone, J.K., Bacon, C.W. & White, J.F. (2000). An overview of endophytic microbes: endophytism defined. In: *Microbial endophyte s*(eds. C.W. Bacon and J.F. White). Marcel Dekker Inc. New York, Basel: 3-29
 11. Saikkonen, K., Faeth, S. H., Helander, M. & Sullivan, T. J. (1998). Fungal Endophytes: A Continuum of Interactions with Host Plants. *Annual review of ecology and systematics*, 29, 319–343.
 12. Strobel, G.A (2002). Rainforest Endophyte and Bioactive Product: *Crit Rev. Biotechnol*, 22(4), 315-33.
 13. Lu, L., He J., Yu X., Li G. & Zhang X. (2006). Studies on isolation and identification of endophytic fungi strain SC13 from harmaceutical plant *Sabina vulgaris* Ant. and metabolites. *Acta Agriculturae Boreali-occidentalis Sinica*. 15, 85-89.
 14. Cou L, Huang J, & Li, J. (2007), Fermentation conditions of *Sinopodophyllum hexandrum* endophytic fungus on production of podophyllotoxin. *Food and Fermentation Industries*.; 33, 28-32
 15. Kour, A., Shawl, AS., Rehman, S., Sultan, P., Qazi, P. H., Suden, P., Khajuria, R. K. & Verma, V. (2008). Isolation and identification of an endophytic strain of *Fusarium oxysporum* producing podophyllotoxin from *Juniperus recurva*. *World Journal of Microbiology and Biotechnology*, 24, 1115-112
 16. Zeng, S., Shao, H. & Zhang, L. (2004): An endophytic fungus producing a substance analogous to podophyllotoxin isolated from *Diphylleiasinensis*. *Journal of Microbiology*, 24, 1-2
 17. Guo, B., Wang, Y., Sun, X. & Tang, K. (2008). Bioactive natural products from endophytes: A review. *Applied biochemistry and microbiology*, 44, 136–142.

18. Eyberger, A.L., Dondapati, R. & Porter, J.R. (2006). Endophyte fungal isolates from *Podophyllum peltatum* produce podophyllotoxin. *Journal of Natural Products*, 69, 1121-1124.
19. Puri, S.C., Nazir, A., Chawla, R., Arora, R., Riyaz-ul-Hasan, S., Amna, T., Ahmed, B., Verma, V., Singh, S., Sagar, R., Sharma, A., Kumar, R., Sharma, R. K. & Qazi, G. N. (2006). The endophytic fungus *Trametes hirsuta* as a novel alternative source of podophyllotoxin and related aryl tetralin ligands. *Journal of Biotechnology*, 122, 494-510.
20. Breinhold, J., Ludvigsen, S. & Rassing, B.R. (1997). Oxysporidinone: a novel antifungal, N-methyl-4-hydroxy-2-pyridinone from *Fusarium oxysporum*. *Journal of Natural Products*, 60, 33-39.
21. Jayasinghe, L., Abbas, H. K. & Jacob, M.R. (2006). A new pentaketide antifungal agent isolated from an endophytic fungus, *Journal of Natural Products*, 69, 439.
22. Khan, A.M., Hamayun, Y-H., Kim, S-M., Kang, J-H. & Lee, I.J (2011). *Process Biochemistry*, 46, 440-447
23. Jalgaonwala, R.E. & Mahajan, R.T. (2011). Evaluation of hydrolytic enzyme activities of endophytes from some indigenous medicinal plants. *Journal of Agricultural Technology*, 7(6), 1733-1741
24. Strobel, G. and Daisy, B. (2003). Bioprospecting for microbial endophytes and their natural products. *Microbiology and molecular biology reviews*, 67, 491-502.
25. Chifu, B.H., Brain, J. & Jeffery, L.E. (2010). Antimicrobial activity of n-6, n-7 and n-9 fatty acids and their esters for oral microorganisms. *Arch Oral Biol.* 55(8), 555-560.
26. Rahmani, N. & Heydarian, Z. (2016). Investigation of *in vitro* antifungal activity of *Salicornia iranica* Akhiani. *Trad Integr Med.* 1(1), 44-46.
27. Chandrasekharan, M., Kannathasan, K. & Venkatesalu, V. (2008). Antimicrobial activity of fatty acid methyl esters of some members of chenopodiaceae. *Z. Natureforsch.* 63C (5-6), 331-6.
28. Gehan, M.A., Hanan, A.E., Hassan, A.H.I., and Okbah, M.A. (2009). Marine natural products and their potential applications as anti-infective agents. *World Sciences Journal*, 7(7), 872-880

29. Muhammad, Y.H., Ghaidaa, J.M. & Imad, H.H. (2016). Analysis of bioactive chemical compounds of *Nigella sativa* using GC-MS. *Journal of Pharmacognosy and Phytotherapy*, 8(2), 8-24
30. Huda, M.A.S (2010). Antimicrobial activity of certain bacteria and fungi isolated from soil mixed with human saliva against pathogenic microbes causing dermatological diseases. *Saudi Journal of Biological Science*, 17, 331–339.
31. Abubakar, U.S., Abdu, G.T., Aliyu I., Fatima, S.S., Jamila, G.A., Tsoken, B. G., Saidu, S. R & Osodi, F.A. (2016). Medicinal Plants Used By Traditional Healer for Treatment of Malaria in Kano Metropholis. *Proceeding of 2nd NWUCIAIC* Nov, 2016. Pp. 138-142.
32. Nishioka, M., Furuya, N., Matsuyama, N. (1997). Antibacterial activity of metabolites produced by *Erwinia* spp. against various phytopathogenic bacteria. *Annu. Phytopathol. Soc. Jap.* 63, 99-102
33. Huang, Y., Wang, J., Li, G., Zheng, Z. & Su, W. (2001). Antitumor and Antifungal activities in endophytic fungi isolated from Pharmaceutical Plant, *Taxus Mairei*, *Cephalataxus fortune* and *Torreya grandis*. *FEMS Immunol. Medi. Microbiol.* 31, 163-167
34. Shrestha, K., G. A. Strobel, S. Prakash & Gewali, M. (2001). Evidence for paclitaxel from three new endophytic fungi of Himalayan yew of Nepal. *Planta Med.* 67, 374-376
35. Harper, J.K., Ford, E.J., Strobel, G.A., Grant, D.M., Porco, J., Tomer D.P. & Oneil, K. (2003). Pestacin A 1,3-dihydro isobenzofuran from *Pestalotiopsis* microspore Possessing antimoxidant and antimycotic activities. *Tetrahedron* 59, 2471-2476